

The overall stereochemical outcome of going from chiral [$^1\text{H}, ^2\text{H}, ^3\text{H}$]- N_5 -methyl tetrahydrofolate to acetate has also recently been determined to be retention,²³ supporting the above sequence.

Acknowledgment. This research was supported by N.I.H. Grants GM 32333 (H.G.F.) and GM 31574 (W.H.O.-J., C.T.W.) and an N.I.H. postdoctoral fellowship (EM 10826) to S.A.R.

(23) Lebertz, H.; Simon, H.; Courtney, L. F.; Benkovic, S. J.; Zydowsky, L. D.; Lee, K.; Floss, H. G. *J. Am. Chem. Soc.*, following paper in this issue.

Stereochemistry of Acetic Acid Formation from 5-Methyltetrahydrofolate by *Clostridium thermoaceticum*

H. Lebertz,[†] H. Simon,*[†] L. F. Courtney,[†] S. J. Benkovic,*[†]
L. D. Zydowsky,[§] K. Lee,[§] and H. G. Floss*[§]

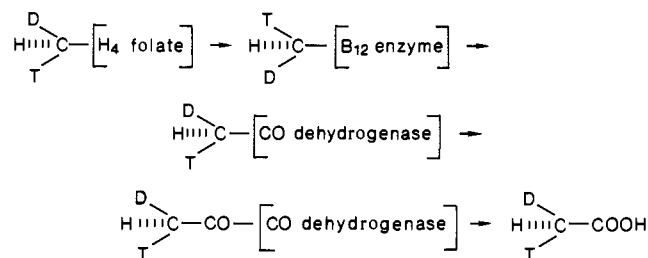
*Lehrstuhl für Organische Chemie und Biochemie der
TU München, D 8046 Garching, West Germany
Department of Chemistry, Pennsylvania State University
College Park, Pennsylvania 16802
Department of Chemistry, The Ohio State University
Columbus, Ohio 43210*

Received December 30, 1986

Some acetogenic bacteria ferment hexoses almost stoichiometrically to 3 mol of acetic acid per 1 mol of sugar.^{1,2} Two of the three moles of acetic acid arise by decarboxylation of pyruvate formed via the Embden-Meyerhof pathway, and the third mole, remarkably, is produced reductively from 2 mol of CO_2 .^{3,4} Thus, an organism like *Clostridium thermoaceticum* can synthesize acetic acid entirely from CO or from $\text{CO}_2 + \text{H}_2$.⁵ One mole of CO_2 is reduced via formate, 5,10-methenyltetrahydrofolate, and 5,10-methylenetetrahydrofolate to 5-methyltetrahydrofolate,^{6,7} and another mole is reduced to CO.⁸ These two products condense in the presence of coenzyme A to produce acetyl-CoA, which then gives acetate with formation of 1 mol of ATP. The formation of acetyl-CoA from methyltetrahydrofolate requires several enzymes, including CO dehydrogenase,^{9,10} a B_{12} enzyme,¹¹⁻¹³ and a methyltetrahydrofolate: B_{12} methyltransferase.^{13,14}

The role of CO dehydrogenase was originally thought to be limited to the reduction of CO_2 to CO,¹³ and it was assumed that the methyl- B_{12} species might give rise directly to the acetyl group,^{8,11-14} by either a carboxylation or a carbonylation of the

Scheme I. Stereochemical Fate of the Methyl Group in the Conversion of Methyltetrahydrofolate to Acetic Acid by *C. thermoaceticum*



methyl-cobalamin. Recent studies by Wood and co-workers, however, point to a far more central role of CO dehydrogenase in this process, as the enzyme on which the actual assembly of acetyl-CoA from CoASH, CO, and a methyl group takes place.⁵ The most critical piece of evidence is the finding that purified CO dehydrogenase alone is capable of catalyzing exchange between CO and the carbonyl group of acetyl-CoA.¹⁵ This exchange requires that the enzyme can bind the methyl, the carbonyl, and the CoA group of acetyl-CoA. Binding of CO to CO dehydrogenase generates a paramagnetic nickel-iron-carbon center,¹⁶ and EPR studies have shown that acetyl-CoA and CoA bind near this center.¹⁵ This suggests the possibility that the acetyl group may be formed by a "carbonyl insertion" reaction on nickel or iron.

It should be possible to test further the mechanism proposed by Wood and co-workers based on stereochemistry. Intermolecular methyl transfers proceed with inversion of configuration.^{17,18} Chemically catalyzed carbonyl insertion, more properly migratory insertion,¹⁹ reactions proceed with retention of alkyl group configuration.²⁰⁻²³ Wood's mechanism involves two methyl transfers, one from methyltetrahydrofolate to B_{12} and another from methyl- B_{12} to CO dehydrogenase, and an insertion reaction; the predicted overall stereochemical outcome would be net retention of methyl group configuration, if retention of configuration is assumed for the CO dehydrogenase catalyzed insertion reaction, too. A process involving carbonylation on the cobalt of B_{12} , on the other hand, would only include one methyl transfer and the insertion reaction, and should thus proceed with net inversion of methyl group configuration.²⁴

To examine this question we incubated (*methyl-R*)- and (*methyl-S*)-[methyl- $^2\text{H}_1, ^3\text{H}$]methyltetrahydrofolate (1.0×10^6 and 1.15×10^5 dpm of ^3H , respectively, in less than $0.1 \mu\text{mol}$), prepared by sequential reduction of 5,10-methenyltetrahydrofolate with deuteriated and tritiated sodium borohydride,²⁶ anaerobically under a CO atmosphere for 30 min at 55°C with a cell-free extract of *C. thermoaceticum* in the presence of 0.1 mM CoASH, 4 mM ATP, 1.4 mM Fe^{2+} , 16 mM dithiothreitol, and 90 mM potassium phosphate, pH 6.0.¹⁴ Following denaturation with HClO_4 , acetic acid was isolated, with carrier dilution, in 17.4% and 23.5% yield, respectively, by passage of the reaction mixture through a column of 8 mL of Dowex 50 H^+ , neutralization and

[†] Lehrstuhl für Organische Chemie und Biochemie der TU München.

[†] Pennsylvania State University.

[§] The Ohio State University.

(1) Fontaine, E. F.; Peterson, D. H.; McCoy, E.; Johnson, M. J.; Ritter, G. J. *J. Bacteriol.* **1942**, *43*, 701.

(2) Ljungdahl, L. G. In *Organic Chemicals from Biomass*; Wise, D. L., Ed.; Benjamin-Cummings: Menlo Park, CA, 1983; pp 219-248.

(3) Barker, H. A.; Kamen, M. D. *Proc. Natl. Acad. Sci. U.S.A.* **1945**, *31*, 219.

(4) Wood, H. G. *J. Biol. Chem.* **1952**, *199*, 579.

(5) For a review, see: Wood, H. G.; Ragsdale, S. W.; Pezacka, E. *Biochem. Int.* **1986**, *12*, 421.

(6) Ljungdahl, L. G.; Wood, H. G. In *Vitamin B₁₂*; Dolphin, D., Ed.; Wiley: New York, 1982; Vol. 2, pp 165-202.

(7) Clark, J. E.; Ljungdahl, L. G. *J. Biol. Chem.* **1984**, *259*, 10845.

(8) Hu, S.-I.; Drake, H. L.; Wood, H. G. *J. Bacteriol.* **1982**, *149*, 440.

(9) Diekert, G. B.; Thauer, R. K. *J. Bacteriol.* **1978**, *136*, 597.

(10) Drake, H. L.; Hu, S.-I.; Wood, H. G. *J. Biol. Chem.* **1980**, *255*, 7174.

(11) Poston, J. M.; Kuratomi, K.; Stadtman, E. R. *Ann. N.Y. Acad. Sci.* **1964**, *112*, 804.

(12) Ljungdahl, L.; Irion, E.; Wood, H. G. *Biochemistry* **1965**, *4*, 2771.

(13) Hu, S.-I.; Pezacka, E.; Wood, H. G. *J. Biol. Chem.* **1984**, *259*, 8892.

(14) Drake, H. L.; Hu, S.-I.; Wood, H. G. *J. Biol. Chem.* **1981**, *256*, 11137.

(15) Ragsdale, S. W.; Wood, H. G. *J. Biol. Chem.* **1985**, *260*, 3970.

(16) Ragsdale, S. W.; Wood, H. G.; Antholine, W. E. *Proc. Natl. Acad. Sci. U.S.A.* **1985**, *62*, 6811.

(17) Arigoni, D. *Ciba Found. Symp.* **1978**, *60*, 243.

(18) Floss, H. G. *ACS Symp. Ser.* **1982**, *185*, 229-243.

(19) Collman, J. P.; Hegedus, L. S. *Principles and Applications of Organotransition Metal Chemistry*; University Science Books: Mill Valley, CA, 1980; pp 260-266.

(20) Bock, P. I.; Boschetto, D. J.; Rasmussen, J. R.; Demers, J. P.; Whitesides, G. M. *J. Am. Chem. Soc.* **1974**, *96*, 2814 and references therein.

(21) Collman, J. P. *Acc. Chem. Res.* **1975**, *8*, 342.

(22) Stille, J. K.; Lau, K. S. Y. *Acc. Chem. Res.* **1977**, *10*, 434.

(23) Walborsky, H. M.; Allen, L. E. *J. Am. Chem. Soc.* **1971**, *93*, 5466.

(24) An earlier attempt²⁵ to examine the steric course of the transformation methyl- $\text{B}_{12} \rightarrow$ acetic acid failed, because the chemically synthesized methyl- B_{12} proved to be racemic.

(25) Imfeld, M. Ph.D. Dissertation No. 6565, ETH, Zürich, 1980.

(26) Zydowsky, T. M.; Courtney, L. F.; Frasca, V.; Kobayashi, K.; Shimizu, H.; Yuen, L. D.; Matthews, R. G.; Benkovic, S. J.; Floss, H. G. *J. Am. Chem. Soc.* **1986**, *108*, 3152.

evaporation of the effluent, and steam distillation of the acidified residue. The acetic acid samples were subjected to chirality analysis by the method of Cornforth *et al.*²⁷ and Arigoni and co-workers,²⁸ using the procedure described by Floss and Tsai.²⁹

The samples of chiral methyl methyltetrahydrofolate had been degraded in earlier work²⁶ and found to contain 44% ee (*R*)-methyl groups and 37% ee (*S*)-methyl groups, respectively. The acetic acid obtained from the (*R*)-methyltetrahydrofolate gave an *F* value¹⁷ of 59.5, corresponding to 33% ee *R* configuration of the methyl group. Analysis of a sample from a second incubation with the same substrate gave *F* equals 58.3% or 29% ee *R* configuration. The acetic acid generated from (*S*)-methyltetrahydrofolate in two independent analyses gave *F* values of 37.2 and 37.2, corresponding to 44% ee *S* configuration of the methyl group.³⁰ It follows that the methyl group of methyltetrahydrofolate is converted by *C. thermoaceticum* into the methyl group of acetic acid with overall retention of configuration. This result argues against acetyl group formation directly from the B₁₂ enzyme but is consistent with the mechanism proposed by Wood and collaborators^{5,15} involving transfer of the methyl group from methyltetrahydrofolate to B₁₂ and then to CO dehydrogenase followed by carbonylation on the latter (Scheme I).

Acknowledgment. We thank the National Institutes of Health (Grants GM 32333 to H.G.F. and GM 24729 to S.J.B.), the Deutsche Forschungsgemeinschaft (SFB 145 to H.S.), and the Alexander v. Humboldt Foundation (Senior U.S. Scientist Award to H.G.F.) for financial support.

(27) Cornforth, J. W.; Redmond, J. W.; Eggerer, H.; Buckel, W.; Gut-schow, C. *Eur. J. Biochem.* **1970**, *14*, 1.

(28) L  thy, J.; R  tey, J.; Arigoni, D. *Nature (London)* **1969**, *221*, 1213.

(29) Floss, H. G.; Tsai, M. D. *Adv. Enzymol.* **1979**, *50*, 243.

(30) *F* values in our assay are accurate to ± 2 ; the *F* values of precursor and product are the same within the limits of accuracy of the assay.

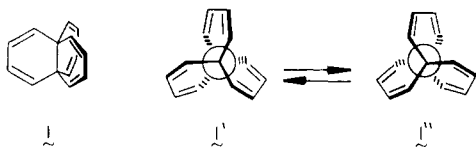
Synthesis of [4.4.4]Propellahexaene

Liladhar Waykole and Leo A. Paquette*

Evans Chemical Laboratories
The Ohio State University, Columbus, Ohio 43210

Received January 20, 1987

Two limitations have severely restricted synthetic access to [4.4.4]propellahexaene (**1**) and its derivatives. Due to proximity



factors, intraring reactions are not easily inhibited.¹ Neither are cationic rearrangements capable of profoundly reorganizing the carbocyclic framework readily circumvented.² One fundamental property of the maximally unsaturated D₃-symmetric hydrocarbon **1** and its derivatives relates to the unusual arrangement of the constituent six-membered rings, which radiate as blades from a common axis (see **1'** \rightleftharpoons **1''**). Extensive theoretical³ and experimental⁴ discussion attests to the general interest attached to this

(1) (a) Paquette, L. A.; Jendralla, H.; DeLuca, G. *J. Am. Chem. Soc.* **1984**, *106*, 1518. (b) Jendralla, H.; Jelich, K.; DeLuca, G.; Paquette, L. A. *Ibid.* **1986**, *108*, 3731.

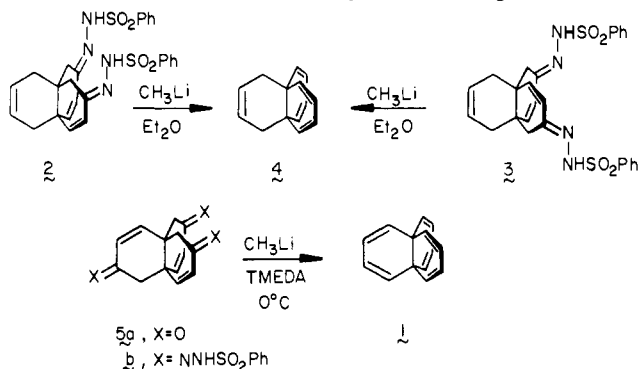
(2) Paquette, L. A.; Waykole, L.; Jendralla, H.; Cottrell, C. E. *J. Am. Chem. Soc.* **1986**, *108*, 3739.

(3) (a) Mislow, K.; Gust, D.; Finocchiaro, P.; Boettcher, R. J. *Top. Curr. Chem.* **1974**, *47*, 1. (b) Farina, M.; Morandi, C. *Tetrahedron* **1974**, *30*, 1819.

dynamical conformation question.

The present purpose is to detail the first synthesis of **1** and to call attention to unprecedented reactions that can materialize upon the attempted preparation of **1** substituted with methoxyl groups. To construct **1**, the pitfalls alluded to above had to be consciously avoided and reactions promising little or no risk of intramolecularly implemented.

In order to evaluate the possible merits of the Shapiro reaction,⁵ a mixture of the bis(phenylsulfonyl)hydrazones **2** and **3** was prepared and treated with methyllithium in ether (0 °C \rightarrow room temperature). Following the identification of **4** as product, the previously described [4.4.4]propellatrienetrione **5a**^{4a} was similarly transformed almost quantitatively into **5b** (mp 195 °C). Base-induced elimination of benzenesulfonic acid and nitrogen from **5b** (CH₃Li, TMEDA, 0 °C) expediently gave the desired **1** (colorless needles, mp 48 °C, 15% isolated). To our knowledge, this reaction sequence represents only the second time a triple-Shapiro degradation has been deployed in a synthetic strategem.⁶



The NMR spectra of **1** reflect its D₃ symmetry. In CDCl₃ at 500 MHz, the vinyl protons appear as an AA'XX' pattern with A centered at δ 5.91 and X at δ 5.27. Its three carbon types resonate in CDCl₃ at 129.44, 122.65, and 34.35 ppm. The electronic spectrum (in isoctane) is characterized by several maxima: 234 (ϵ 25 730), 242 (24 100), 251 (27 780), 271 (9560), 279 (7615), and 291 nm (6975).

The hexaene is remarkably stable in air. After heating bromobenzene-*d*₃ solutions of **1** at 105 °C for 4.5 h, the ¹H NMR spectrum remained unaltered. An increase in temperature to 150 °C caused gradual decomposition to unidentified nonaromatic products (*t*_{1/2} \approx 5 h). We cannot exclude the possibility that this destruction was catalyzed by traces of acid.

In an attempt to produce **9**, triketone **6**^{4a} was treated sequentially with excess potassium *tert*-butoxide in dry DMF and dimethyl sulfate,⁷ all at 0 °C. Chromatography of the resulting mixture on basic alumina afforded the dimethoxy pentaenone **7** (31%),⁸ 2,7-dimethoxynaphthalene (**8**, 3.4%), and trace quantities of **9**, which were visible only in the ¹H NMR spectrum of the unpurified product. Complex mixtures were obtained when **7** was resubmitted to the original reaction conditions or to the action of KN(SiMe₃)₂/THF-Me₂SO₄ at low temperatures. In contrast, the combination of KH in anhydrous DMF (0 °C) and Me₂SO₄ gave rise to **8** and **9** in a 17:1 ratio (¹H NMR analysis).⁹ Since

(4) (a) Jendralla, H.; Doecke, C. W.; Paquette, L. A. *J. Chem. Soc., Chem. Commun.* **1984**, 942. (b) Gilboa, H.; Altman, J.; Loewenstein, A. *J. Am. Chem. Soc.* **1969**, *91*, 6062. (c) Ermer, O.; Gerdil, R.; Dunitz, J. D. *Helv. Chim. Acta* **1971**, *54*, 2476. (d) Alder, R. W.; Arrowsmith, R. J.; Bryce, M. R.; Eastment, P.; Orpen, A. G. *J. Chem. Soc., Perkin Trans. 2* **1983**, 1519.

(5) Shapiro, R. H. *Org. React.* **1976**, *23*, 405.

(6) Font, J.; Lopez, F.; Serratos, F. *Tetrahedron Lett.* **1972**, 2589.

(7) Paquette, L. A.; Broadhurst, M. J. *J. Org. Chem.* **1973**, *38*, 1886.

(8) ¹H NMR (300 MHz, CDCl₃) δ 6.27 (d, *J* = 10 Hz, 1 H), 6.02 (d, *J* = 10 Hz, 1 H), 5.92 (dd, *J* = 9.7, 2.1 Hz, 2 H), 5.51 (d, *J* = 9.7 Hz, 2 H), 4.29 (d, *J* = 2.1 Hz, 2 H), 3.51 (s, 6 H), 2.48 (s, 2 H); ¹³C NMR (75 MHz, CDCl₃) ppm 199.23, 152.33, 144.32, 128.85, 127.64, 124.50, 99.45, 54.38, 46.87, 42.13, 41.08; MS, *m/z* (*M*⁺) calcd 256.1099, obsd 256.1109.

(9) (a) The IR and ¹H NMR spectra of **8** (mp 137-139 °C) were identical with those of a commercial sample (Aldrich). (b) For **9**: ¹H NMR (300 MHz, C₆D₆) δ 5.99 (dd, *J* = 9.8, 2.1 Hz, 3 H), 5.19 (d, *J* = 9.8 Hz, 3 H), 4.35 (d, *J* = 2.1 Hz, 3 H), 3.23 (s, 9 H); MS, *m/z* (*M*⁺) calcd 270.1256, obsd 270.1235.